

# Dosimetry and methodology of gamma irradiation for degradation studies on solvent extraction systems

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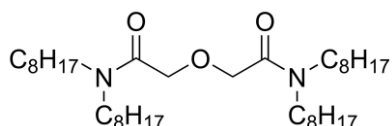
## Abstract

The recycling of minor actinides from dissolved nuclear fuels by hydrometallurgical separation is one challenging strategy for the management of spent fuel. These future separation processes will likely be based on solvent extraction processes in which an organic solvent system (extractant and diluent) will be contacted with highly radioactive aqueous solutions. To establish a separation between different elements in spent nuclear fuel, many extractants have been studied in the past. A particular example is *N,N,N',N'*-tetraoctyl diglycolamide (TODGA), which co-extracts lanthanides and actinides from nitric acid solutions into an organic phase (*e.g.* TODGA in *n*-dodecane). The radiolytic stability of these extractants is crucial, since they will absorb high doses of ionizing radiation during their usage. Worldwide, different gamma irradiation facilities are employed to expose extractants to ionizing radiation and gain insight in their radiation stability. The facilities differ in many ways, such as their environment (pool-type or dry), configuration and gamma sources (often <sup>60</sup>Co or spent nuclear fuel). In this paper, a dosimetric assessment is made using different dosimeter systems in a pool-type irradiation facility, which has the advantage to be flexible in its arrangement of <sup>60</sup>Co sources. It is shown that Red Perspex dosimeters can be used to accurately characterize this high dose rate gamma irradiation field (*approx.* 13.6 kGy h<sup>-1</sup>), after comparison with alanine, Fricke and ceric-cerous dosimetry in a lower dose rate gamma irradiation field (*approx.* 0.5 kGy h<sup>-1</sup>). A final validation

31 of the whole chain of techniques is obtained by reproduction of the dose constants for TODGA  
32 in *n*-dodecane.

## 33 1 Introduction

34 Waste produced by nuclear power plants remains highly radiotoxic for millions of years.  
35 Therefore, safe management of long-lived radioactive waste including final long-term storage  
36 of the spent nuclear fuel poses a challenge. In spent nuclear fuel, plutonium and the minor  
37 actinides are the main contributors of radiotoxicity and heat production on the long term. The  
38 size of the necessary final repository is mainly defined by this heat production.[1] To minimize  
39 the space required in the deep geological repository, new separation processes for actinides  
40 based on liquid-liquid extractions are developed. Here, a separation is obtained by adding an  
41 organic phase (*e.g.* kerosene) with complexing agents to dissolved spent fuel solutions. Initially,  
42 the Plutonium Uranium Reduction EXtraction (PUREX) process was developed to recover the  
43 major actinides (U and Pu) from spent fuel.[2] In the following decades, more research has been  
44 conducted to also recover the minor actinides[3-4] in order to further decrease the long-term  
45 activity of the residual spent fuel inventory, which led to developments of new complexing  
46 agents or ligands, such as diglycolamides.[5-6] The most important, and up to date, most  
47 intensely studied member of this class of ligands is *N,N,N',N'*-tetraoctyl diglycolamide  
48 (TODGA) of which the structure is shown in Figure 1. As the solvents used in these  
49 hydrometallurgical processes need to be operated in contact with radioactive feed solutions, it  
50 is crucial to not only evaluate new compounds on their extraction behavior, but to also study  
51 how they are affected by ionizing irradiation.



53 *Figure 1: Structure of TODGA.*

54 Radioactive decay causes degradation of the organic phase in different ways. Ionizing radiation  
55 can directly cause disintegration of certain bonds of ligand molecules. However, studies on  
56 radiolysis of TODGA have shown that its degradation is dominated by reactions with radical  
57 cations of the diluent.[7-8]

58 For the comparison of the radiolytic degradation of different ligands or to study the effect of  
59 changing the diluent, dose constants (*d*) are being used in this research.[9] The dose constant is  
60 determined by the slope of the linear fit of the natural logarithm of the concentration of the  
61 molecule as a function of the absorbed dose. This slope can be considered as a constant if the  
62 degradation exhibits (pseudo) first-order kinetics. The use of the *G* and *G<sub>o</sub>* values in this context  
63 was discussed by Mincher *et al.*[9] The main weakness of the use of *G* values is that it depends

64 on the concentration, which changes in the case of continuous irradiation experiments. For this  
65 reason it could be more useful to define  $G_o$ , which is defined as the slope of the concentration  
66 as a function of the dose, but only over a very small interval near the initial concentration.[10]  
67 In this case,  $G_o$  is the slope of a line tangent to the curve and it can be expressed as the dose  
68 constant multiplied with the initial concentration and converted to the appropriate units ( $C_o$  is  
69 the initial concentration in  $\mu\text{mol kg}^{-1}$ ) using equation (1).[9, 11] For simplicity and consistency,  
70 in this work only dose constants will be compared.

$$G_o = d \times C_o \quad (1)$$

71 The ligand TODGA is a well-studied molecule which is known to co-extract actinides and  
72 lanthanides efficiently due to the tridentate coordination through hard-donor oxygen atoms.[12-  
73 14] Its radiolysis behavior has been studied intensively,[15-18] which makes it a usable  
74 reference for further radiolysis research of solvent extraction systems.

75 The gamma sources in previous radiolysis stability studies were either  $^{60}\text{Co}$  sources,[15, 19-20]  
76 or spent nuclear fuel sources.[21-22] The gamma spectrum of the latter is much broader than  
77 the spectrum of  $^{60}\text{Co}$  because of the presence of different fission and activation products such  
78 as  $^{137}\text{Cs}$ ,  $^{134}\text{Cs}$ ,  $^{154}\text{Eu}$  and  $^{60}\text{Co}$ . Furthermore, Bremsstrahlung, which is generated by the  
79 deceleration of electrons and positrons in the Coulomb field of nuclei, contributes significantly  
80 to the total photon source spectrum of a spent nuclear fuel source. It should be noted that there  
81 is also a difference between different  $^{60}\text{Co}$  sources. Gamma irradiations are either conducted  
82 using dry facilities[15, 17-18, 20, 23-25] (*e.g.* Gammacell 220), or pool-type facilities[16, 19,  
83 26-27] (*e.g.* Náyade at CIEMAT). In the latter, the spectrum will be degraded because of  
84 passing through water and steel.[28] In general, gamma irradiation facilities are used for many  
85 purposes: modification of polymers,[29] sterilization in medical applications,[30-31] food  
86 preservation[32] and for radiolysis studies on solvents used for reprocessing of spent nuclear  
87 fuels.[7, 15-20, 23, 33-38] Also for testing material resistance in general, these facilities prove  
88 to be very useful, *e.g.* for testing of fiber-optics for space industry.[39]

89 In an experimental pool-type  $^{60}\text{Co}$  gamma ray field used for irradiation studies, the complexity  
90 of the variable irradiation geometry (number, configuration and activity of the loaded  $^{60}\text{Co}$   
91 sources) requires a dosimetric assessment in advance of any experiment. Schematic  
92 representations of these irradiation set-ups are shown in Figure 2. It is a great advantage that  
93 sources can be added and repositioned to obtain the best possible conditions for certain  
94 experiments, although this makes characterization of the irradiation field crucial. The selection

95 of suitable dosimetry systems that was used for the assessment, was made on the basis of  
96 international recommendations, reviews and standards elaborated for high dose rate field  
97 radiation processing facilities.[40-42] Alanine,[41] Fricke[43] and ceric–cerous[44] dosimeters  
98 are classified as reference-standard dosimeters (type I), in contrast to Perspex dosimeters which  
99 are classified as routine (type II) dosimeters.[41] The advantage of type I dosimeters is that they  
100 are of high metrological quality: different quantifiable influences are very well identified and  
101 can be compensated by using correction factors. However, application of these involves many  
102 manipulations and handling of chemicals, which consume a lot of time and resources. On the  
103 other hand, routine type II dosimeters, such as Perspex dosimeters, are easy to use and do not  
104 require any preparations, except precise positioning. The disadvantage of this type of  
105 dosimeters is that the response is influenced in a complex way for which it is not possible to  
106 apply individual correction factors.[41] Responses may vary because of environmental  
107 conditions such as temperature, light, relative humidity and the presence of gases.[45] The use  
108 of alanine and Fricke dosimetry is limited to low dose rate facilities (in our case  $\pm 0.5 \text{ kGy h}^{-1}$ ),  
109 since absorbed doses during positioning in high dose rate fields would get too significant for  
110 these sensitive dosimeters. An overview of the applicable working ranges (absorbed doses) of  
111 all applied dosimeter systems is shown in Figure 3. Here, it can be easily noticed that alanine  
112 and Fricke dosimeters are indeed on the low side of the absorbed dose ranges compared to  
113 Perspex and ceric–cerous dosimeters. The advantage of ceric–cerous and Fricke dosimetry  
114 systems over the others, is that these dosimeters can be tailored to the actual size of the sample  
115 to be irradiated, so that the exact geometrical position occupied by the sample to be irradiated  
116 can be accurately assessed. Also, these liquid dosimeters have similar absorption properties  
117 with respect to gamma radiation as the samples which will be studied (extractant dissolved in  
118 an organic diluent). The homogeneity of the used irradiation field is evaluated by mapping the  
119 local dose rates of the different sample positions in the sample holder. This information is  
120 crucial, since it will be a great asset to use this kind of flexible sources in which the  
121 configuration of the sources can be easily modified to the needs of the user.

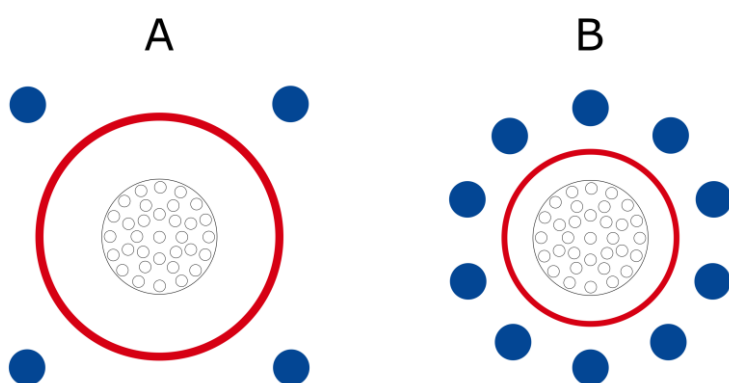
122 In this work, the dose constants of TODGA in *n*-dodecane were determined using a pool-type  
123  $^{60}\text{Co}$  gamma irradiation facility and validated against literature values achieved with dry  $^{60}\text{Co}$   
124 sources. The validation of our irradiation methodology posed several challenges. The first  
125 challenge was the validation of different dosimeters and the transfer of dose rate calibration  
126 from low dose rate gamma fields to high dose rate gamma fields. In this paper, the dosimetric  
127 assessment of such a flexible, high dose rate irradiation facility is described in detail. Different

128 type I dosimeters, of which one is read-out externally, were used to conduct this comparison.  
129 The use of various independent methods was chosen in this study to reveal any systematic error  
130 that could otherwise remain unnoticed with respect to the dose calibration. Another challenge  
131 in this study is the high dose rate gradient due to the close distance to the sources. The final  
132 validation of the irradiation set-up for radiolysis studies of solvent extractions systems was  
133 performed *via* chemical analysis of an irradiated well-characterized solvent (TODGA in *n*-  
134 dodecane). This study opens perspectives for future research using flexible, high dose rate pool-  
135 type gamma irradiation facilities on novel solvent extraction systems, such as newly developed  
136 diglycolamides[19, 46] or ionic-liquid-based systems.[47-49]

## 137 2 Experimental

### 138 2.1 Irradiation facilities

139 The gamma irradiations were conducted in BRIGITTE (Big Radius Installation under Gamma  
140 Irradiation for Tailoring and Testing Experiments) and RITA (Radio Isotope Test Arrangement)  
141 of SCK CEN (Mol, Belgium).[50] Here, underwater (pool-type)  $^{60}\text{Co}$  sources ( $E_{\gamma 1} = 1173.2$  keV  
142 and  $E_{\gamma 2} = 1332.5$  keV) are used at an approximate dose rate of respectively  $13.6$  kGy  $\text{h}^{-1}$   
143 (BRIGITTE) and  $0.510$  kGy  $\text{h}^{-1}$  (RITA). These  $^{60}\text{Co}$  sources are old control rods of the Belgian  
144 Reactor 2 (BR2), where  $^{59}\text{Co}$  parts were activated during reactor operation. Schematic  
145 representations of the set-up in RITA and BRIGITTE are shown in Figure 2.



146

147 *Figure 2: A: Schematic representation of the irradiation container (red circle) with an inserted sample holder between four*  
148  *$^{60}\text{Co}$  sources (filled blue circles) in RITA (top view). B: Schematic representation of the irradiation container (red circle) with*  
149 *an inserted sample holder between ten  $^{60}\text{Co}$  sources (filled blue circles) in BRIGITTE (top view)*

### 150 2.2 Dosimeters

151 Glassware used for the preparation of the Fricke and ceric-cerous dosimeter solutions and for  
152 irradiation was cleaned according to the following protocol adapted from the relevant  
153 standard.[43] Two stock solutions of  $0.4$  mol  $\text{L}^{-1}$  and  $4$  mol  $\text{L}^{-1}$   $\text{H}_2\text{SO}_4$  (Optima grade, Fisher  
154 Scientific UK Limited, Loughborough) in MilliQ water (obtained from a Sartorius Arium pro  
155 UV instrument with a built-in UV-digester and Total Organic Carbon (TOC) analyzer; TOC <  
156  $5$  ppb, specific resistance:  $18.2$   $\text{M}\Omega$  cm) were prepared in  $1$  L glass volumetric flasks. The  
157 prepared  $4$  mol  $\text{L}^{-1}$   $\text{H}_2\text{SO}_4$  stock solution was irradiated in BRIGITTE to  $500$  Gy absorbed dose  
158 in order to eliminate traces of organic contaminants in the water and sulfuric acid, which  
159 possibly has an effect on the ferrous ions.[43] Glass ampoules (Kimble, Illinois USA) of  $5$  mL  
160 volume were rinsed twice with MilliQ water (TOC <  $5$  ppb), followed by filling with MilliQ  
161 water and boiling for  $1$  h on a hot plate. The ampoules were emptied and dried in an oven at  
162  $100$   $^\circ\text{C}$  until dry, followed by baking in a furnace at  $550$   $^\circ\text{C}$  for  $1$  h. The ampoules were filled

163 with the dosimetry solutions using glass Pasteur pipettes and flame sealed using a  
164 propane/oxygen gas mixture.

165 Due to the extreme sensitivity of the method to trace impurities only new glassware was used  
166 for the preparation of the stock solutions and intermediate dilutions. The glassware necessary  
167 for the dilutions was cleaned by ultrasound with MilliQ water at 60 °C for 1 h, followed by  
168 rinsing with hot concentrated sulfuric acid. The sulfuric acid was washed thoroughly away with  
169 MilliQ water and the glassware was dried in an oven at 100 °C.

170 The reported errors on the average of the determined dose rates with alanine, Fricke and ceric-  
171 cerous dosimetry are the 99 % confidence intervals as a result of using standard deviations of  
172 multiple measurements and t-values corresponding to the relevant degrees of freedom. For  
173 Perspex dosimetry, this interval was determined by multiplication of the propagated uncertainty  
174 (based on the reported measurement reproducibility and calibration quality by the  
175 manufacturer) and a factor of 2.58 to obtain similar 99 % confidence intervals. Confidence  
176 intervals were only reported in Table 1, in other cases standard deviations are used to express  
177 error bars.

### 178 **2.2.1 Alanine dosimetry**

179 Alanine dosimeters (air-tight sealed plastic foil packages each containing 5 alanine pellets of  
180 5 mm diameter and 2.5 mm thickness), were received from NuTec (Hasselt, Belgium), and  
181 irradiated in a PMMA grid up to 5, 10, 15, 20 and 25 Gy absorbed dose in the RITA facility at  
182 a height of 35 cm, in the center of the irradiation container. The irradiated, unopened packages  
183 were returned to NuTec for read-out by Electron Paramagnetic Resonance Spectroscopy  
184 (EPR)[51], using a Bruker EMXmicro spectrometer with a 9 inch magnet, equipped with a  
185 high-sensitivity resonator ER4119HS-W1, following a protocol described by Anton *et al.*[52-  
186 53]

### 187 **2.2.2 Fricke dosimetry**

188 The Fricke dosimeters were prepared and used in accordance with the ISO/ASTM standard  
189 51026:2015 on the Standard Practice for Using the Fricke Dosimetry System.[43] The stock  
190 solutions for the dosimeters were prepared by adding 98 mg of  $(\text{NH}_4)_2\text{Fe}(\text{SO}_4)_2 \cdot 6\text{H}_2\text{O}$   
191 (99.997 %, Sigma Aldrich, Overijse, Belgium) and 14.5 mg of NaCl (99.999 % Sigma Aldrich,  
192 Overijse, Belgium), to suppress any effects of possible carbon contaminations, to a 250 mL  
193 volumetric flask. The salts were dissolved in 100 mL of the 0.4 mol L<sup>-1</sup> H<sub>2</sub>SO<sub>4</sub> stock solution.

194 After dissolution, the volume was adjusted using 0.4 mol L<sup>-1</sup> H<sub>2</sub>SO<sub>4</sub>. For the calculation of the  
195 absorbed doses, temperature corrections were made as described in the ASTM standard.

196 Absorbed doses were calculated using equation (2).

$$D = \frac{\Delta A}{G_{T_{irrad}} \times \varepsilon_{T_{meas}} \times \rho \times d} \quad (2)$$

197 In this equation  $\varepsilon_{25}$  and  $G_{25}$  are corrected for measurement and irradiation temperature by  
198 equations (3) and (4), with  $\varepsilon_{25} = 1.61 \times 10^{-6}$  mol J<sup>-1</sup> and  $G_{25} = 1.61 \times 10^{-6}$  mol J<sup>-1</sup>.

$$\varepsilon_{T_{meas}} = \varepsilon_{25}[1 + 0.0069(T_{meas} - 25)] \quad (3)$$

$$G_{T_{irrad}} = G_{25}[1 + 0.0012(T_{irrad} - 25)] \quad (4)$$

### 199 **2.2.3 Ceric–cerous dosimetry**

200 Both high and low range ceric–cerous dosimeters (abbreviated as CCH and CCL, respectively)  
201 were prepared in accordance with the ISO/ASTM51205:2009 standard.[44] For the preparation  
202 of the Ce<sup>4+</sup> stock solution, 58.0006 g of Ce(SO<sub>4</sub>)<sub>2</sub>·4H<sub>2</sub>O (Optima grade, Fisher Scientific UK  
203 Limited, Loughborough) was dissolved in 1 L stock solution of 0.4 mol L<sup>-1</sup> H<sub>2</sub>SO<sub>4</sub>. A Ce<sup>3+</sup>  
204 stock solution was prepared by dissolving 31.0061 g of Ce<sub>2</sub>(SO<sub>4</sub>)<sub>3</sub>·8H<sub>2</sub>O (99.999 %, Sigma  
205 Aldrich, Overijse, Belgium) in a total volume of 1 L MilliQ water. Both stock solutions were  
206 allowed to stand at least two weeks in the dark to stabilize in case of the presence of any carbon  
207 contaminations. For the preparation of the low range dosimeter solution, 15 mL of the Ce<sup>3+</sup>  
208 stock solution, 15 mL of the Ce<sup>4+</sup> stock solution and 48.5 mL of 4 mol L<sup>-1</sup> H<sub>2</sub>SO<sub>4</sub> were  
209 combined in a volumetric flask and filled up with MilliQ water up to a total volume of 500 mL.  
210 The high range dosimeter solution was prepared by combining 75 mL of the Ce<sup>3+</sup> stock solution,  
211 75 mL of the Ce<sup>4+</sup> stock solution and 42.5 mL of 4 mol L<sup>-1</sup> H<sub>2</sub>SO<sub>4</sub> in a volumetric flask, which  
212 was further filled with MilliQ water to a total volume of 500 mL. Both dosimetry solutions  
213 were allowed to stand for at least five days in the dark, as recommended.[44] Changes in  
214 absorbance ( $\Delta A$ ), induced by irradiation, are converted into absorbed doses ( $D$ ) by using  
215 equation (5).

$$D = \frac{f \times \Delta A}{G(Ce^{3+}) \times \varepsilon_m \times \rho \times d} \quad (5)$$

216 In this equation,  $f$  is the dilution factor for the irradiated dosimeters,  $G(Ce^{3+})$  is calculated as  
217 mentioned in the in the ASTM (equation (6)),  $\varepsilon_m$  is the molar-linear absorption coefficient

218  $(561 \text{ m}^2 \text{ mol}^{-1})$ ,  $\rho$  is the density of the dosimetric solution ( $1.032 \times 10^3 \text{ kg m}^{-3}$ ) and  $d$  is the path  
219 length of the spectrophotometric cell (0.010 m).

$$\begin{aligned} G(\text{Ce}^{3+})(\text{low range}) &= (2.42452 - 0.0052 \times T) \times 1.036 \times 10^{-7} \\ G(\text{Ce}^{3+})(\text{high range}) &= (2.33544 - 0.0052 \times T) \times 1.036 \times 10^{-7} \end{aligned} \quad (6)$$

#### 220 **2.2.4 Perspex (PMMA) dosimetry**

221 The dose calibration of the irradiation chambers was performed using either Harwell's Red  
222 Perspex dosimeters (Type 4034,  $\lambda = 640 \text{ nm}$ ) or Amber Perspex dosimeters (Type 3042,  $\lambda =$   
223  $603 \text{ nm}$  or  $651 \text{ nm}$ ) (Harwell Dosimetrics, Oxfordshire, UK). The 3 mm thick slits were placed  
224 in a UV-VIS cell holder (Hellma Analytics BVBA, Krubek, Belgium) to ensure a  
225 reproducible positioning and perpendicular alignment with respect to the light beam. The  
226 absorbance of the dosimeters belonging to a given batch was converted to absorbed dose using  
227 the calibration curve determined by Harwell. Harwell's calibration is traceable to the UK  
228 standard of absorbed dose at the National Physical Laboratory. For Amber Perspex the relative  
229 standard deviation of specific absorbance measurements on sets of dosimeters is 2.5 % and for  
230 Red Perspex, it is 2 %.

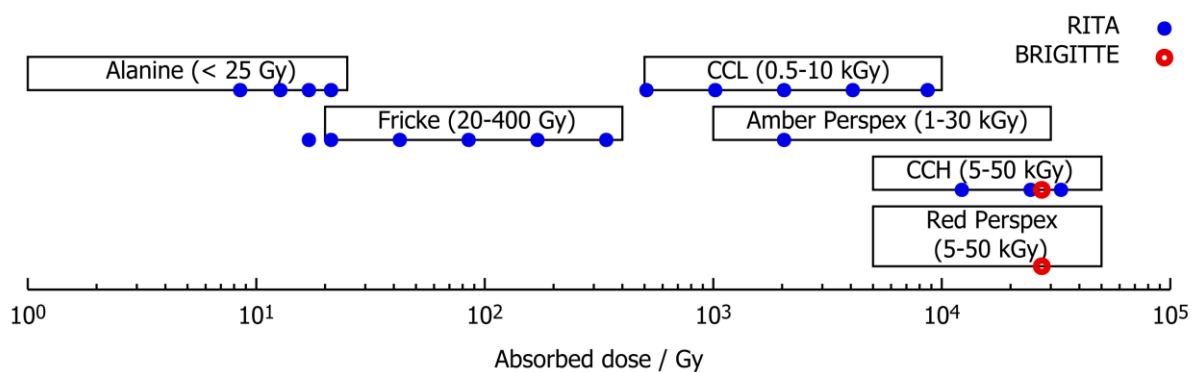
#### 231 **2.3 UV-VIS spectrometry**

232 The working principle of the Fricke, ceric-cerous and Perspex dosimeters is based on the  
233 change in absorption of the dosimeter material under the effect of absorbed gamma radiation.  
234 The UV-VIS measurements were performed using a Shimadzu UV-1800 spectrophotometer  
235 with a spectral bandwidth of 1 nm. The spectrophotometer performance was verified after  
236 baseline correction by the use of certified reference materials obtained from Hellma Analytics  
237 BVBA, following the relevant ASTM standard.[54] The accuracy of the wavelength scale in  
238 both the ultraviolet and visible region of the electromagnetic spectrum was verified against a  
239 holmium oxide glass filter 666-F1 and a holmium perchlorate liquid filter 667-UV-5 certified  
240 reference (Hellma Analytics, certified by Deutsche Akkreditierungsstelle GmbH). The  
241 absorbance scale accuracy of the instrument in the visible region was verified to be within the  
242 95 % confidence intervals reported on the certificate with the use of F2, F3 and F4 neutral  
243 density glass filters (Hellma Analytics, all certified by Deutsche Akkreditierungsstelle GmbH)  
244 at 440.0 nm, 465.0 nm, 546.1 nm, 546.1 nm, 590.0 nm and 635.0 nm. The absorbance accuracy  
245 of the spectrophotometer in the UV-region was verified by the use of liquid reference materials  
246 667 UV-20, 667 UV-60, and UV-600 against UV-14 in the reference cell (Hellma Analytics,  
247 certified by Deutsche Akkreditierungsstelle GmbH).

248 The read-out wavelength for Fricke dosimetry was 303 nm, ceric-cerous dosimeters were read-  
 249 out at 320 nm. Red Perspex dosimeters' absorbances were determined at 640 nm and Amber  
 250 Perspex dosimeters at 603 nm. For all liquids, cuvettes with a path length of 10 mm were used.

## 251 2.4 Irradiation of dosimeters

252 All dosimeters were irradiated in RITA and/or BRIGITTE. The Amber variant of Perspex  
 253 dosimeters was loaded in RITA, whereas in BRIGITTE only ceric-cerous and  
 254 Red Perspex dosimeters were irradiated. An overview of the employed dosimeters, delivered  
 255 doses and irradiation facilities is shown in Figure 3. Also, the dose range per dosimeter is  
 256 indicated in this schematic.



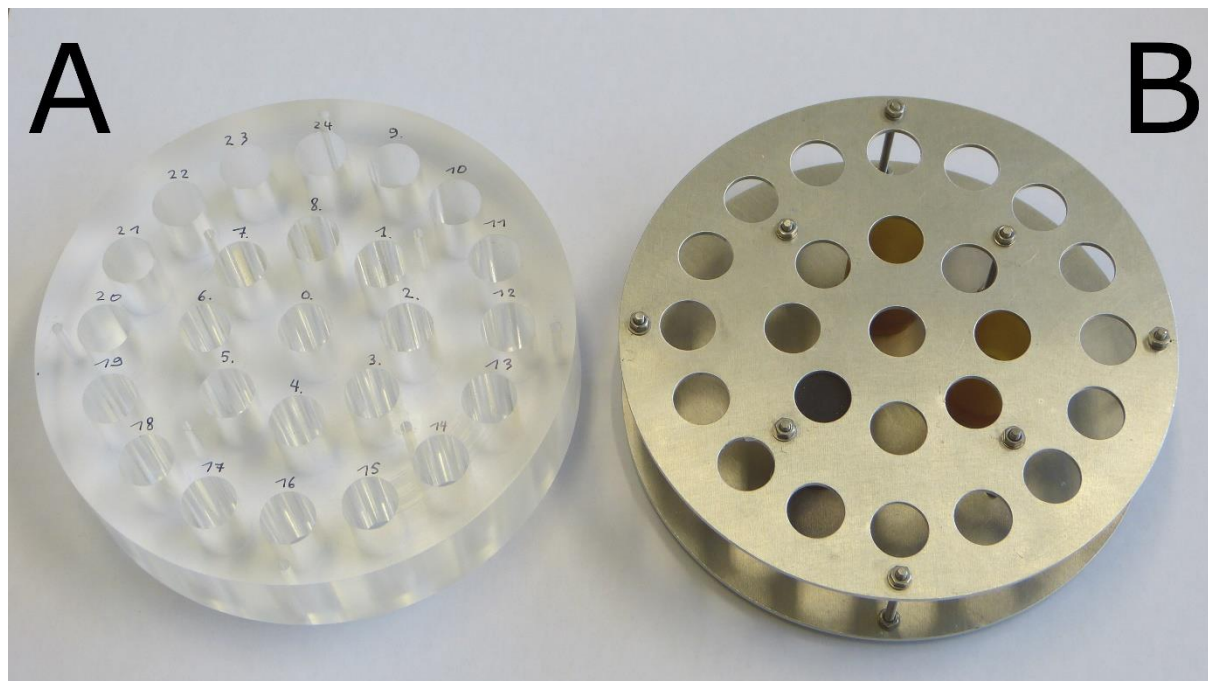
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258 *Figure 3: Schematic of the working ranges of all applied dosimeters, with delivered doses by RITA (blue closed dots) and by*  
 259 *BRIGITTE (red open dots). Low range ceric-cerous dosimeters and high range ceric-cerous dosimeters are abbreviated as*  
 260 *CCL and CCH.*

261 For the irradiation of dosimeters and samples, in-house manufactured poly(methyl  
 262 methacrylate) (PMMA) sample holders were used, similar to the one shown in Figure 4. The  
 263 choice of the material of the sample holder (PMMA) was made in order to establish secondary  
 264 charged particle equilibrium in the sample positions since the thickness of PMMA is more than  
 265 3 mm.[55] All positions are numbered for dose rate mapping. The high dose rate gamma field  
 266 of BRIGITTE was characterized by placing the same PMMA sample holder as used in RITA  
 267 with its central position aligned to the central axis of the cylinder-shaped irradiation container,  
 268 at 110 cm distance from the lid of the container.

269 Experiments with an aluminum sample holder and epoxy resin cups (thickness = 3.77 mm)  
 270 were conducted with Red Perspex dosimeters in BRIGITTE. The metal parts were put together  
 271 as shown in Figure 4. This holder has a diameter of 160 mm and is 35 mm high, holes for  
 272 positioning samples have a diameter of 16 mm. The aluminum holder will ensure stable  
 273 placement of the samples, even if the polymer degrades due to the high absorbed dose, and

274 makes it possible to study the effect of the environment in which the Red Perspex dosimeters  
275 were irradiated.



276

277 *Figure 4: PMMA sample holder (A) and aluminum sample holder in which epoxy cups are inserted (B) for gamma irradiations*  
278 *in BRIGITTE.*

## 279 **2.5 Radiolysis study of TODGA**

280 A solution of 0.05 mol L<sup>-1</sup> TODGA (Technocomm Ltd., Wellbrae United Kingdom) in *n*-  
281 dodecane (Lot 16C020503, VWR) was prepared; 0.5 mL fractions were transferred to 1.5 mL  
282 glass vials for irradiation. To half of the samples, equal volumes of 2.5 mol L<sup>-1</sup> trace metal grade  
283 HNO<sub>3</sub> solution (Fisher Scientific, Merelbeke, Belgium) were added.

284 Gamma irradiations were conducted in BRIGITTE in the PMMA sample holder which was  
285 used for dosimetry. The absorbed doses were 100, 200, 300 and 400 kGy, based on dosimetry  
286 using the Red Perspex dosimeters. A sample for aging control was used as the reference.

287 Quantitative HPLC-ESI-MS/MS was performed with a Qtrap6500 instrument (ABSciex,  
288 Darmstadt, Germany) coupled by electrospray ionization with an Agilent 1260 HPLC. The used  
289 column was of the type Phenyl-X (100 × 4.6 mm; 2.6 μm) (Thermo Fisher). Elution was  
290 obtained with a gradient of H<sub>2</sub>O with 0.1 % formic acid and acetonitrile with 0.1 % formic acid,  
291 at a flow rate of 0.7 mL min<sup>-1</sup>. The temperature of the column oven was set at 40 °C.  
292 Quantification after HPLC separation was performed using ESI-MS/MS detection in the  
293 multiple reaction-monitoring (MRM) mode, with 581.4 *m/z* as the parent ion and 312.2 *m/z* and  
294 340.2 *m/z* as the product ions.

295 **3 Results and discussion**

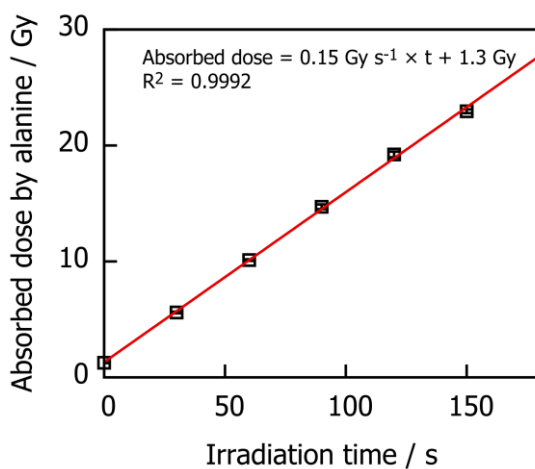
296 **3.1 Irradiation of dosimeters in RITA**

297 The actual absorbed dose rate in the center position in the irradiation cavity of RITA was  
298 determined using different dosimetric methods. Samples and dosimeters were at the maximum  
299 dose rate and in the axially homogeneous part of the irradiation field. In the span [-25 mm, +25  
300 mm], the dose rate variation is less than 2 %. The height of both the samples and dosimeters  
301 (approx. 30 mm) was well within this homogeneous zone The value of the ‘transient dose’, *i.e.*  
302 the absorbed dose which the samples receive during positioning, loading and unloading, was  
303 obtained from alanine dosimetry. The transient dose is used to make a correction on the read-  
304 out values of the dosimeters in RITA.

305 **3.1.1 Alanine dosimeters**

306 The results of the consecutive dosimetry in the central position in RITA using alanine  
307 dosimeters with increasing irradiation times are shown in Figure 5. The transient dose, with 0 s  
308 residence time) was  $1.26 \pm 0.02$  Gy. This agrees with the y-axis intercept ( $1.3 \pm 0.2$  Gy) of the  
309 linear fitting of the response as a function of the irradiation time. The value of 0.9992 for  $R^2$  of  
310 the linear least square fitting, indicates that the dependency of the read-out of the absorbed dose  
311 is linear to the irradiation time.

312 The average dose rate calculated from the five consecutively irradiated, transient dose  
313 corrected, packages of 5 alanine pellets is  $530 \pm 18$  Gy h<sup>-1</sup>. The determined dose rates as a  
314 function of irradiation time are shown in Figure 6. Alanine dosimetry was conducted to obtain  
315 an independent (read-out externally) traceable dose calibration with a Type I dosimeter as a  
316 calibration transfer dosimeter.

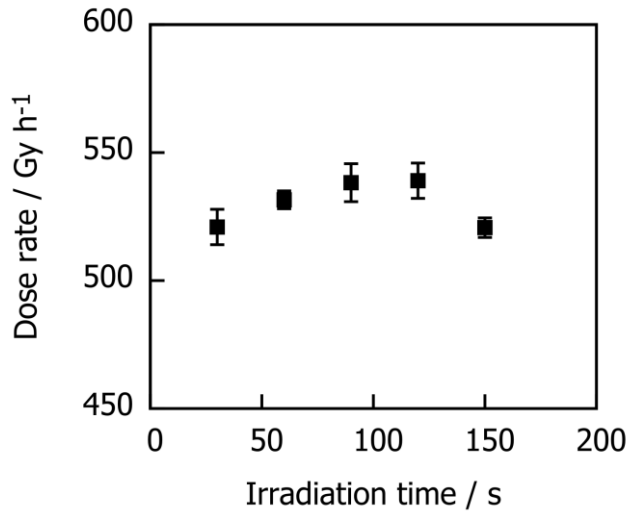


317

318 *Figure 5: Absorbed doses determined by alanine/EPR dosimetry as a function of irradiation time. The error bars represent*  
319 *standard deviations of the 5 repetitions in every dosimeter package.*

320

321



322

323 *Figure 6: Dose rate in RITA determined with alanine/EPR dosimetry. The error bars represent standard deviations of the 5*  
324 *repetitions in every dosimeter package.*

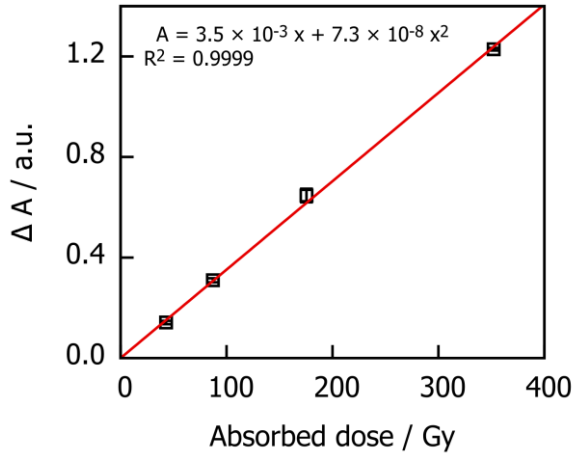
### 325 **3.1.2 Fricke dosimeters**

326 The Fricke dosimeters were used to confirm the dose rate values obtained from the alanine  
327 dosimeters. Fricke dosimeters are liquid type I dosimeters, which are prepared in sample-sized  
328 ampoules (as described in 2.2.2). In Figure 7, the calibration curve of the Fricke dosimeters in  
329 RITA is shown. The change in absorbance can be fitted to a second order polynomial function  
330 of the absorbed dose as shown in equation (7).[43]

$$\Delta A = b_0 + b_1 \times D + b_2 \times D^2 \quad (7)$$

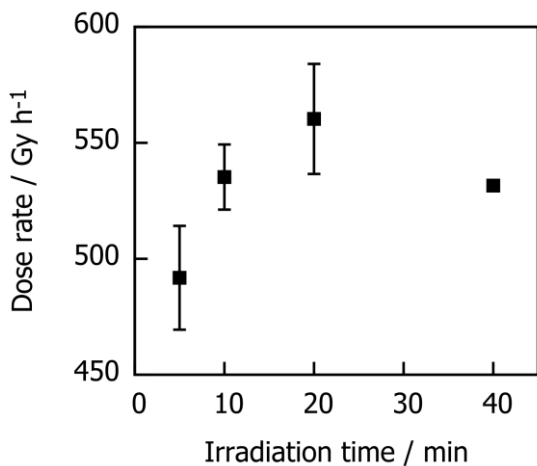
331 In this equation is  $\Delta A$  the change in absorbance,  $D$  the absorbed dose and  $b_0$ ,  $b_1$  and  $b_2$  are the  
332 coefficients of the second order polynomial fit. The value of coefficient  $b_1$  equals  
333  $(4.0 \pm 0.3) \times 10^{-3}$ , when  $b_0$  is considered to be zero  $b_1$  equals  $(3.5 \pm 0.1) \times 10^{-3}$  ( $R^2 = 0.9999$ ).  
334 The latter assumption is legitimated by the fact that no change in absorbance is assumed if the  
335 dosimeters are not irradiated, the  $R^2$  of 0.9999 confirms this. This result of  $b_1$  is comparable to  
336 the value reported in the ASTM for Fricke dosimetry[43] of  $3.6 \times 10^{-3}$ , confirming a good match  
337 between the dose rate determined with Fricke dosimeters and the alanine/EPR methods. The  
338 coefficient for the second order part of the equation is relatively small, which means that the  
339 curve is very close to being linear. The reason for this term, is that towards the end of Fricke's  
340 application range this second order part can become more significant.

341 In Figure 8 the determined dose rate is shown as a function of the irradiation time, all the  
 342 absorbed doses were in the working range of the Fricke dosimeters. The average determined  
 343 dose rate is  $530 \pm 27 \text{ Gy h}^{-1}$ .



344

345 *Figure 7: Calibration curve of Fricke dosimeters in RITA, error bars represent the standard deviation of three repeated*  
 346 *measurements. The change in measured absorbance is visualized as function of the absorbed dose, determined by alanine*  
 347 *dosimetry.*

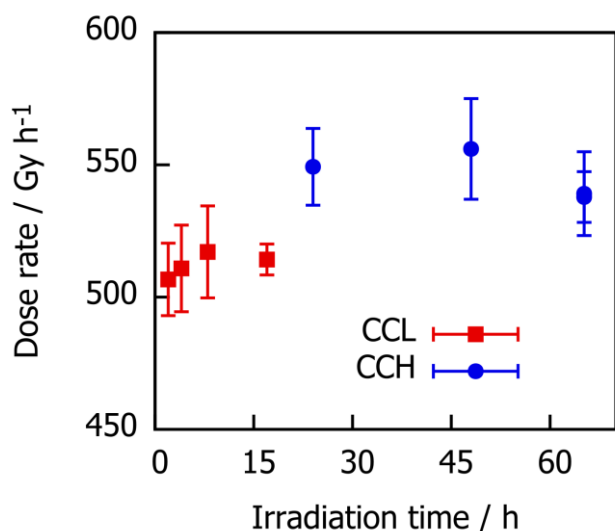


348

349 *Figure 8: Dose rate in RITA determined with Fricke dosimetry using the literature values for Fricke dosimetry. Every data*  
 350 *point is based on three repetitions, standard deviations on these repetitions are visualized by the error bars.*

### 351 **3.1.3 Ceric–cerous dosimetry**

352 Determined dose rates for different irradiation times with low range ceric–cerous dosimeters  
 353 are shown in Figure 9. The average determined dose rate was  $513 \pm 17 \text{ Gy h}^{-1}$ . High range  
 354 ceric–cerous dosimeters were also irradiated for three different durations. The average dose rate  
 355 at this central position of the cavity was  $546 \pm 14 \text{ Gy h}^{-1}$ .



356

357 *Figure 9: Dose rate determined with CCL (red squares) and CCH (blue dots). Every irradiation was repeated three times, of*  
 358 *which the standard deviation was calculated and visualized by error bars.*

### 359 **3.1.4 Perspex dosimeters**

360 The Perspex dosimeters (Amber and Red) are routine dosimeters of Type II classification under  
 361 the ISO/ASTM standard on the practice for dosimetry in radiation processing.[41] Their ease  
 362 of use (in comparison to ceric-cerous dosimeters) make them a very practical tool to perform  
 363 routine dosimetry and particularly for dose rate mapping.[56] The determined dose rate based  
 364 on the specific absorbance of the irradiated Amber Perspex dosimeters at the center of the  
 365 sample holder was  $510 \pm 33 \text{ Gy h}^{-1}$ . This was derived from the calibration curve provided by  
 366 the supplier of the dosimeters. By using Amber Perspex dosimeters in RITA and using Red  
 367 Perspex dosimeters in BRIGITTE, the irradiation time (2 h) is kept identical because of their  
 368 difference in application range, thus eliminating possible effects caused by positioning.

### 369 **3.1.5 Overview of dosimetry in RITA**

370 Table 1 lists the dose rate values determined with all the dosimetry methods used in this study.  
 371 The shown errors are the 99 % confidence intervals, as a result of the repeated measurements.  
 372 None of the reported values are significantly different. The dose rates determined with CCL  
 373 and Amber Perspex seem a bit low and CCH a bit high compared to Fricke and alanine  
 374 dosimetry, although confidence intervals still overlap in these cases. This is crucial information,  
 375 since alanine and Fricke dosimeters are not practically usable in the high dose rate field of  
 376 BRIGITTE (*approx.*  $13.6 \text{ kGy h}^{-1}$ ). The alanine and Fricke dosimeters would be saturated  
 377 within less than a few minutes, and the dose they receive during emplacement of the irradiation  
 378 canister would become very significant. For further experiments concerning dose rate mapping  
 379 in a higher irradiation field, both ceric–cerous high range and Red Perspex dosimeters are used.

380 *Table 1: Comparison of dose rate in RITA at 35 cm height, central position determined with the various dosimeter systems.*

Method	Determined dose rate (Gy h <sup>-1</sup> )	Reference
Alanine	530 ± 18	ISO/ASTM 51607[57]
Fricke	530 ± 27	ISO/ASTM 51026[43]
Ceric–cerous low range	513 ± 17	ASTM 51205[44]
Ceric–cerous high range	546 ± 14	ASTM 51205[44]
Amber Perspex	510 ± 33	ISO 51276[56]

381

## 382 **3.2 Irradiation of dosimeters in BRIGITTE**

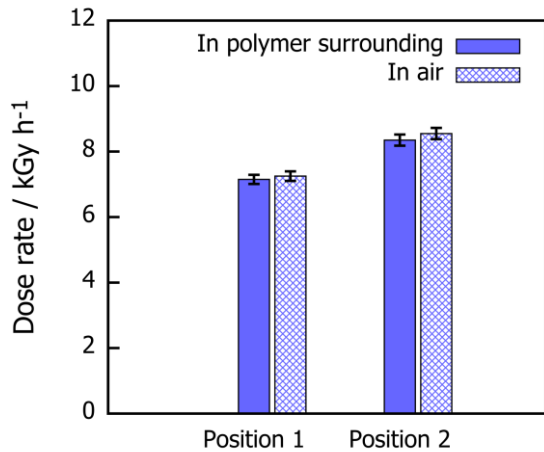
### 383 **3.2.1 Influence of the sample holder**

384 One of the reasons to use the previously mentioned PMMA block, is reproducible positioning  
 385 of samples. However, since polymers degrade under gamma irradiation and finally lose all  
 386 structural integrity as the absorbed dose accumulates, using a metal sample holder would be a  
 387 great improvement for gamma irradiations in a high dose rate gamma irradiation facility such  
 388 as BRIGITTE. For this purpose, an aluminum holder was designed.

389 While this seems like the perfect solution, dosimeters are not properly surrounded to ensure a  
 390 charged particle equilibrium. For this reason, a separate experiment was set-up, in which the  
 391 determined dose rate with and without a polymer (epoxy resin) shielding were compared. The  
 392 thickness of the epoxy cups, of 3.77 mm, should ensure a charged particle equilibrium,[55]  
 393 without causing large shielding effect.

394 The data shown in Figure 10, indicate there is no significant difference between irradiation of  
 395 Red Perspex dosimeters in the epoxy resin containers and the plain dosimeters in the aluminum  
 396 holder for the two positions. Reproducible dosimetry with Perspex dosimeters is possible, even  
 397 without using a polymer surrounding. For the two control positions (were the direct  
 398 environment of the dosimeter was not changed), there was also no significant difference  
 399 observed; control 1:  $8.70 \pm 0.17 \text{ kGy h}^{-1}$  and  $8.70 \pm 0.17 \text{ kGy h}^{-1}$ , control 2:  $7.55 \pm 0.15 \text{ kGy h}^{-1}$   
 400 <sup>1</sup> and  $7.45 \pm 0.15 \text{ kGy h}^{-1}$ .

401 For future experiments, the aluminum sample holder can thus be used with no need to insert a  
 402 PMMA block for conducting dosimetry and following irradiations. This is important, since in  
 403 high irradiation fields, such as BRIGITTE, these kind of blocks are destroyed during  
 404 irradiations over a few MGy, which was observed in practice.

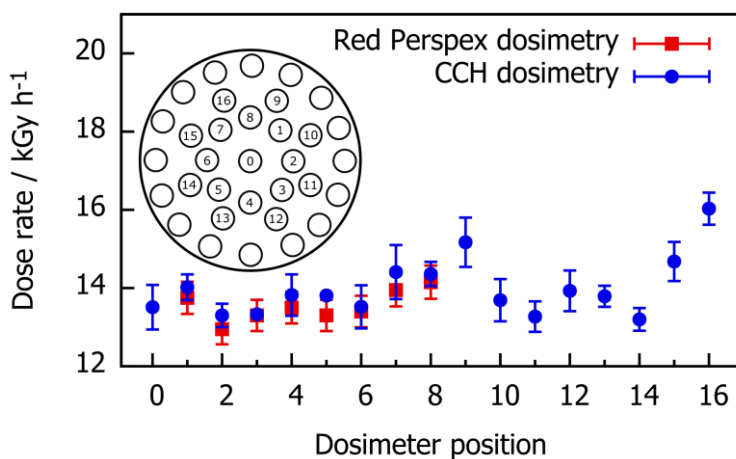


405

406 *Figure 10: Determined dose rates with Red Perspex dosimetry in the aluminum sample holder with (solid fill) and without*  
 407 *(checkered fill) epoxy resin shielding.*

408 **3.2.2 Dose rate mapping**

409 The BRIGITTE irradiation facility uses activated control rods of the BR2 reactor, installed  
 410 centro-symmetrically around the BRIGITTE container. The use of a uniform dose rate  
 411 throughout a series of samples is necessary to exclude effects related to differences in dose  
 412 rates. An analysis of the dose rate distribution in space was performed by using CCH and Red  
 413 Perspex dosimeters placed in the central axis of the cylinder-shaped irradiation cavity and at  
 414 the perimeter of a 10 cm diameter circle. The result of this dose rate mapping is visualized in  
 415 Figure 11, error bars for Red Perspex are 2 %, as this is the coefficient of variation reported by  
 416 the manufacturer. For the high range ceric–cerous dosimeters standard deviations of three  
 417 repetitions are shown.



418

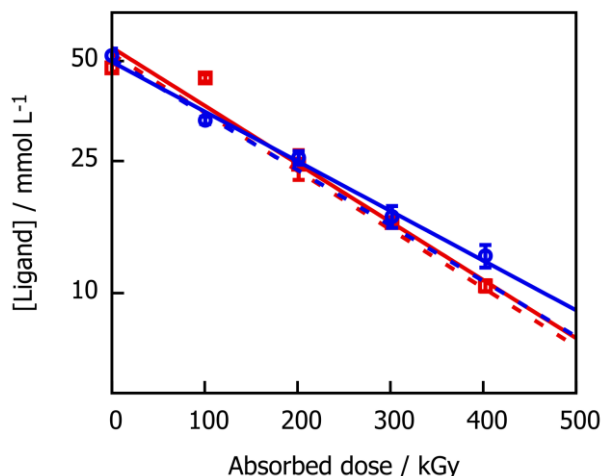
419 *Figure 11: Dose rate mapping using high range ceric–cerous and Red Perspex dosimeters in a PMMA sample holder in*  
 420 *BRIGITTE. The inset shows the positioning of samples in the sample holder.*

421 The sample-size matched dose rate mapping for the samples was performed by the use of high  
422 range ceric–cerous dosimeters in the PMMA sample holder block. The delivered dose to the  
423 solvent extraction samples was calculated by multiplication of the irradiation time with the  
424 average dose rate (determined with CCH) in the most homogeneous part of the irradiation field.

425 As the dose rate mapping revealed, for three positions the dose rate was higher than the average  
426 by 12 %, 9 % and 19 % respectively (positions 9, 15 and 16). This azimuthal inhomogeneity is  
427 caused by differences in activity of the  $^{60}\text{Co}$  sources. Therefore, these positions were not used  
428 for the irradiation of TODGA samples. By restricting the use to only 14 positions, the applied  
429 dose rate varied with less than 7 % compared to the central position of the sample holder. The  
430 outer positions were omitted since they are closer to the  $^{60}\text{Co}$  gamma sources. Consequentially,  
431 the non-uniformity in the dose rate field will be amplified for these positions, which is  
432 unfavorable for dosimetric studies. Differences in dose rates applied to samples are minimized  
433 by limiting the sample positions to the center, excluding an additional parameter which possibly  
434 influences the behavior of certain radiolysis processes. This influence has been reported during  
435 the irradiation of solvent extraction systems,[58] gold solutions[59] and methyl orange  
436 solutions.[60] The average dose rate in this part of the field is  $13.6 \pm 0.5 \text{ kGy h}^{-1}$ , this is the  
437 value which was used to calculate the absorbed dose of the irradiated TODGA samples in the  
438 next section. Differences in activity of the  $^{60}\text{Co}$  sources cause azimuthal inhomogeneities of the  
439 irradiation field in the cavity. In case all sources have an identical activity, the dose rate  
440 distribution would be symmetric as seen in Gammacell 220 irradiator devices.[61]

### 441 **3.3 Gamma irradiation of TODGA**

442 The ligand concentration as a function of the absorbed dose is plotted in Figure 12. Data shown  
443 in red refers to TODGA in *n*-dodecane, whereas data in blue refers to TODGA in contact with  
444  $2.5 \text{ mol L}^{-1} \text{ HNO}_3$ . Dotted lines are literature values[15] for both TODGA in *n*-dodecane (red)  
445 and TODGA in contact  $2.5 \text{ mol L}^{-1} \text{ HNO}_3$  (blue). Dose constants ( $d$ ) are calculated from the  
446 linear fit of the natural logarithm of the ligand concentration as a function of the absorbed dose.  
447 The irradiations were conducted in the same absorbed dose range with a similar dose rate and  
448 identical gamma energies as mentioned in the literature.[15]



449

450 *Figure 12: Influence of the absorbed dose on the concentration of TODGA in n-dodecane (□), and for TODGA in n-*  
 451 *dodecane in contact with 2.5 mol L<sup>-1</sup> HNO<sub>3</sub> (○) with their respective fittings (—) and (—). Dose constants found by Zarzana*  
 452 *et al.[15] for TODGA in n-dodecane, and TODGA in n-dodecane in contact with 2.5 mol L<sup>-1</sup> HNO<sub>3</sub> are displayed with dotted*  
 453 *lines (- - -) and (- - -).*

454 The determined dose constant for TODGA in *n*-dodecane is  $(4.0 \pm 0.4) \times 10^{-6}$  Gy, which is  
 455 within experimental uncertainty identical to the value reported by Zarzana *et al.*:  $(4.1 \pm 0.3) \times$   
 456  $10^{-6}$  Gy.[15] Also the value for TODGA in HNO<sub>3</sub>,  $(3.4 \pm 0.2) \times 10^{-6}$  Gy, is very similar to the  
 457 literature value of  $(3.8 \pm 0.3) \times 10^{-6}$  Gy.[15] It should be noted that these values from Zarzana  
 458 *et al.* were determined after irradiation with a (dry) <sup>60</sup>Co Gamma Cell source (Nordion, Ottawa,  
 459 Canada).[15] Also dose constants published by Sugo *et al.*[17] are similar ( $4.5 \times 10^{-6}$  Gy) for  
 460 an initial concentration of 0.05 mol L<sup>-1</sup> TODGA in *n*-dodecane, in absence of an aqueous phase.  
 461 These results confirm the reproducibility of the whole chain from sample preparation over  
 462 dosimetry to analysis.

463

#### 464 **4 Conclusions**

465 In this study it is shown that the dose rate constants for TODGA obtained with dry  $^{60}\text{Co}$   
466 irradiation facilities can be reproduced in a pool-type irradiation facility with a more complex  
467 geometry. This is achieved by reproducible sample positioning, validation of dosimetry and  
468 accurate dose rate mapping of the irradiation chamber. It can be concluded that the performance  
469 of Amber Perspex dosimeters is not significantly different from ceric–cerous (high and low  
470 range), Fricke and alanine dosimeters. The latter are type I dosimeters, which are dosimeters of  
471 high metrological quality and for which independent correction factors can be used to express  
472 individual influence quantities in a well-defined way.[41] The results were first confirmed by  
473 irradiations in RITA, the low dose rate facility of *approx.*  $0.5\text{ kGy h}^{-1}$ . Additionally, irradiation  
474 of high-range ceric–cerous and Red Perspex dosimeters in BRIGITTE (the high dose rate  
475 facility, *approx.*  $13.6\text{ kGy h}^{-1}$ ) led to similar results. In future experiments, Red Perspex  
476 dosimetry can be a valid alternative for dosimetry and dose rate mapping of the irradiation  
477 cavity prior to irradiation experiments. Dose rate mapping proved to be crucial for evaluation  
478 of the irradiation field and is therefore strongly advised to be performed whenever the  
479 configuration of the sources is changed or samples are arranged differently. The main  
480 advantages of using Perspex dosimeters for this purpose are: (1) it does not require any  
481 manipulation of chemicals, (2) has a robust read-out method and (3) is applicable to the high  
482 dose rate field of BRIGITTE. Moreover, it is shown that future irradiations could also be  
483 conducted in an aluminum sample holder, without using a PMMA body (which degrades  
484 quickly under high gamma irradiation), since there was no significant effect of the presence of  
485 the polymer on Red Perspex dosimeters observed. Our findings open perspectives to future  
486 radiolysis research using flexible, high dose rate pool-type gamma irradiation facilities.

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